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Ghenia Bentabed-Ababsa, Ely Sidaty Cheikh Sid, Stéphanie Hesse, Ekhlass Nassar, Floris Chevallier, et al.. Direct metalation of heteroaromatic esters and nitriles using a mixed lithium-cadmium base. Subsequent conversion to dipyridopyrimidinones.. Journal of Organic Chemistry, 2010, 75 (3), pp.839-847. 10.1021/jo902385h . hal-00785069

HAL Id: hal-00785069

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Direct metalation of heteroaromatic esters and nitriles using a mixed lithium-cadmium base. Subsequent conversion to dipyridopyrimidinones

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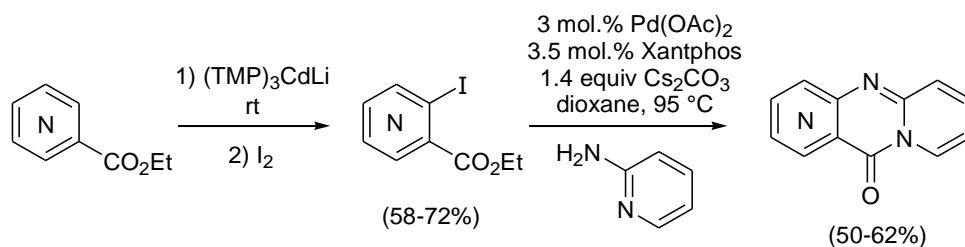
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Abstract:

All pyridine nitriles and esters were metalated at the position next to the directing group using $(\text{TMP})_3\text{CdLi}$ in tetrahydrofuran at room temperature. 2-, 3-, and 4-Cyanopyridine were treated with 0.5 equivalent of base for 2 h to afford, after subsequent trapping with iodine, the corresponding 3-iodo, 2-iodo, and 3-iodo derivatives, respectively, in yields ranging from 30 to 61%. Cyanopyrazine was similarly functionalized at the 3 position in 43% yield. Ethyl 3-iodopicolinate and -isonicotinate were synthesized from the corresponding pyridine esters in 58 and 65% yield. Less stable ethyl 4-iodonicotinate also formed under the same conditions, and was directly converted to ethyl 4-(pyrazol-1-yl)nicotinate in a two steps 38% yield. All three ethyl iodopyridinecarboxylates were involved in a one pot palladium-catalyzed cross-coupling reaction/cyclization using 2-aminopyridine to afford new dipyrrolic compounds: dipyrrolic[1,2- α :3',2'- d]pyrimidin-11-one, dipyrrolic[1,2- α :4',3'- d]pyrimidin-11-one and dipyrrolic[1,2- α :3',4'- d]pyrimidin-5-one in yields ranging from 50 to 62%. A similar cross-coupling/cyclization sequence was applied to methyl 2-chloronicotinate using 2-aminopyridine, 2-amino-5-methylpyridine and 1-aminoisoquinoline to give the corresponding tricyclic or tetracyclic compounds in 43-79% yield. Dipyrrolic[1,2- α :4',3'- d]pyrimidin-11-one and dipyrrolic[1,2- α :3',4'- d]pyrimidin-5-one showed a good bactericidal activity against *Pseudomonas aeruginosa*. Dipyrrolic[1,2- α :2',3'- d]pyrimidin-5-one and pyrido[2',3':4,5]pyrimidino[2,1- α]isoquinolin-8-one showed a fungicidal activity against *Fusarium*, and dipyrrolic[1,2- α :4',3'- d]pyrimidin-11-one against *Candida albicans*. Ethyl 4-(pyrazol-1-yl)nicotinate and dipyrrolic[1,2- α :2',3'- d]pyrimidin-5-one have promising cytotoxic activities, the former toward a liver carcinoma cell line (HEPG2) and the latter toward a human breast carcinoma cell line (MCF7).

Introduction

Interest in pyridine natural products and pharmaceuticals, as well as pyridine building blocks for various applications such as material science, has resulted in extensive efforts on synthesis methodologies.¹ The deprotonative metalation using lithium bases has been widely used as a powerful method for the regioselective functionalization of such substrates.² Nevertheless, the incompatibility of lithium compounds with reactive functions or sensitive heterocycles can be a limit to their use for the elaboration of complex molecules. Recourse to softer magnesium bases can improve the chemoselectivity of deprotonation reactions, but it is to the detriment of their efficiency since a large excess of base has in general to be used.³

The use of metal additives to get more efficient or more chemoselective bases (synergic superbases) has been respectively developed by Schlosser⁴ and Lochmann⁵ with LIC-KOR, mixture of butyllithium (LIC) and potassium *tert*-butoxide (KOR), and by Caubère,⁶ Gros and Fort⁷ in the pyridine series with BuLi-LiDMAE (DMAE = 2-dimethylaminoethoxide). More recently, other $(R)_n(R')_nMLi$ type bases, but with M different from an alkali metal, have been described by different groups.⁸ By combining alkali additives with soft organometallic compounds, bases such as $R_2Zn(TMP)Li(\cdot TMEDA)$ ($R = ^iBu, Bu$; TMP = 2,2,6,6-tetramethylpiperidino) (described by the groups of Kondo, Uchiyama, Mulvey and Hevia),⁹ $(TMP)_2Zn \cdot 2MgCl_2 \cdot 2LiCl$ ¹⁰ and $TMPZnCl \cdot LiCl$ ¹¹ (Knochel), $^iBu_3Al(TMP)Li$ (Uchiyama and Mulvey),¹² $Al(TMP)_3 \cdot 3LiCl$ (Knochel),¹³ $(Me_3SiCH_2)_2Mn(TMP)Li \cdot TMEDA$ (Mulvey),¹⁴ and $MeCu(TMP)(CN)Li_2$ (Uchiyama and Wheatley)¹⁵ have been prepared, characterized and used to generate functionalized aromatic compounds.

We recently accomplished the room temperature deprotonative metalation of a large range of substrates including sensitive heterocycles and functionalized benzenes using a newly developed lithium-cadmium base, $(TMP)_3CdLi$, prepared from $CdCl_2 \cdot TMEDA$ and 3 equivalents of $LiTMP$.¹⁶ If TMEDA is often employed in solvents of low or modest polarities to enhance the reactivity of a base^{9d,e} or to

obtain a specific regioselectivity,² it was here rather used in order to simplify the reaction protocol, CdCl₂·TMEDA being much less sensitive to hydration than free CdCl₂.¹⁷

We here describe the use of (TMP)₃CdLi for the functionalization of pyridine esters and nitriles. Ethyl iodopyridinecarboxylates thus obtained appeared as useful key synthetic intermediates for the synthesis of polycyclic compounds containing a dipyridopyrimidinone skeleton. Some compounds were evaluated for their antimicrobial and cytotoxic activity.

Results and Discussion

Synthetic aspects

Due to their electrophilic functional group and to their ring prone to nucleophilic attacks, cyanopyridines have never been metalated at room temperature. Reactions using cyano as a group to direct *ortho*-lithiation have been reported in the benzene series from 1982,¹⁸ but the first example in the pyridine series only appeared 20 years later. Larock and coll. showed in 2002 that it was possible to lithiate 3-cyanopyridine using LiTMP in tetrahydrofuran (THF) at –78 °C. This result was evidenced by subsequent trapping with iodine to afford a 1:1 mixture of the 2- and 4-iodo compounds in a 50% total yield.¹⁹ Rault and coll. achieved in 2005 the regioselective²⁰ functionalization of the other cyanopyridine isomers using 2 equivalents of the same hindered lithium amide in THF at –80 °C for 0.75 h.²¹

The deprotonation of cyanopyridines (**1-3**) as well as cyanopyrazine (**4**) was attempted using (TMP)₃CdLi in THF (Table 1), this base being suitable to metalate benzonitrile.^{16a} Conducting the reaction from 2-cyanopyridine (**1**) using 0.5 equivalent of base at 0 °C for 2 h resulted, after quenching with iodine, in the formation of a mixture from which the main compound, 2-cyano-3-iodopyridine (**5a**), was isolated in 39% yield (Entry 1). When the reaction was carried out at room temperature, the iodide **5a** formed in 30% yield, due to the more important formation of side products (Entry 2). By using 1 equivalent of base at room temperature, the di- and triiodide **5b,c** were obtained in 28 and 20% yield, respectively (Entry 3). If the formation of a diiodinated compound can be rationalized as the

result of a dimetalation, $(\text{TMP})_3\text{CdLi}$ being able to dideprotonate substrates such as pyrazine,^{16b} thiazole,^{16a} *N*-Boc pyrrole,^{16a} thiophenes^{16a} and [1,2,3]triazolo[1,5-*a*]pyridines,^{16c} the triiodide **5c** could rather result from a metalation of **5b** during the trapping step with iodine, as already suggested in the case of 3-(2-pyridyl)-[1,2,3]triazolo[1,5-*a*]pyridine.^{16c}

The reaction from 4-cyanopyridine (**2**) was then attempted using 0.5 equivalent of base at 0 °C for 2 h; subsequent trapping with iodine afforded a mixture of 4-cyano-3-iodo- and 4-cyano-3,5-diiodopyridine (**6a,b**) in 30 and 20% yield, respectively (Entry 4). By performing the reaction at room temperature, the diiodide was not observed, but a 72:28 ratio of 4-cyano-3-iodopyridine (**6a**) and isomeric 4-cyano-2-iodopyridine (**6c**) was obtained instead, the latter being isolated in 44 and 10% yield, respectively (Entry 5). Surprisingly, carrying out the reaction with 1 equivalent of base resulted in the formation of the diiodide **6d** under the same conditions (Entry 6).

The results obtained with 3-cyanopyridine (**3**) were less disappointing. Indeed, when exposed to 0.5 equivalent of base at room temperature for 2 h, this substrate was regioselectively metalated at the 2 position. This was demonstrated by subsequent interception with iodine to afford the derivative **7** in 61% yield (Entry 7). This regioselectivity is different to that previously documented by other teams using LiTMP in THF at low temperatures; indeed, by using the lithium amide, the metalation took place unregioselectively at the positions adjacent to the cyano group.^{19,21a} Such a result could be partly explained by the presence of a different directing group for the metalation using LiTMP than for that using $(\text{TMP})_3\text{CdLi}$; whereas a first equivalent of LiTMP adds to the cyano group in the study performed by Rault and coll., it does not seem to be the case with $(\text{TMP})_3\text{CdLi}$ (Scheme 1).

SCHEME 1. 3-Cyanopyridine (3): Comparisons of the Species Before Ring Deprotonation using LiTMP and $(\text{TMP})_3\text{CdLi}$.

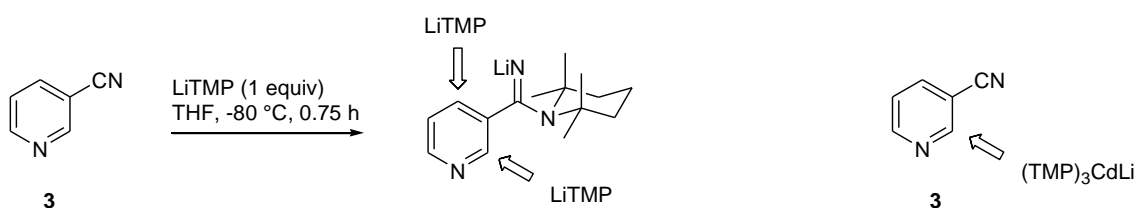


TABLE 1. Deprotonation of 1-4 using (TMP)₃CdLi Followed by Trapping with I₂.

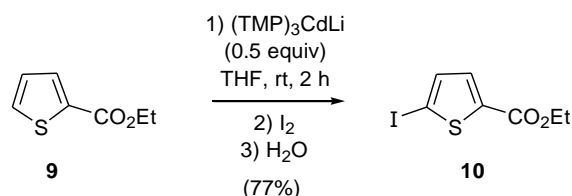
<div style="display: flex; align-items: center; justify-content: center;"> <div style="text-align: center;"> </div> <div style="margin-left: 20px;"> <p>1) (TMP)₃CdLi (x equiv) THF temp., 2 h</p> <p>2) I₂ 3) H₂O</p> </div> <div style="text-align: center;"> <p>5-8</p> </div> </div> <div style="margin-top: 10px;"> <p>1: 2-cyanopyridine 2: 4-cyanopyridine 3: 3-cyanopyridine 4: cyanopyrazine</p> </div>			
entry	substrate	x, temp.	product(s), yield(s)
1	1	0.5, 0 °C	<p>5a, 39%^a</p>
2	1	0.5, rt	5a , 30% ^b
3	1	1, rt	<p>5b, 28%</p> <p>5c, 20%</p>
4	2	0.5, 0 °C	<p>6a, 30%</p> <p>6b, 20%</p>
5	2	0.5, rt	6a , 44%
6	2	1, rt	<p>6c, 10%</p> <p>6d, 51%</p>
7	3	0.5, rt	<p>7, 61%</p>
8	4	0.5, rt	<p>8, 43%^c</p>

^a Other compounds including 2-cyano-3,4-diiodopyridine and 2-cyano-3,6-diiodopyridine were identified in the crude. ^b Other compounds including 2-cyano-6-iodopyridine and 2-cyano-3,6-diiodopyridine were identified in the crude. ^c A mixture of **8** and an unidentified diiodide was obtained in a 75:25 ratio.

These conditions were extended to cyanopyrazine (**4**) for which metalation mainly took place at the position next to the cyano group to furnish the iodide **8** in 43% yield (Entry 8).

The compatibility of an ester function with $(\text{TMP})_3\text{CdLi}$ in THF at room temperature has been recently evidenced with the possible metalation of methyl benzoate.^{16a} Using ethyl thiophene-2-carboxylate (**9**) as substrate also resulted in its cadmation.²² After a 2 h contact with 0.5 equivalent of base followed by quenching with iodine, the 5-iodo derivative **10** was obtained in 77% yield (Scheme 2).

SCHEME 2. Functionalization of Ethyl Thiophene-2-carboxylate (9**) using $(\text{TMP})_3\text{CdLi}$.**



Deprotonation of ethyl pyridinecarboxylates is a much more difficult challenge due to easy nucleophilic attacks on their ring. In 2007, Knochel and coll. reported the magnesiation of ethyl isonicotinate using $(\text{TMP})_2\text{Mg}\cdot 2\text{LiCl}$ in THF at $-40\text{ }^\circ\text{C}$ for 12 h to give, after trapping with iodine, the corresponding 3-iodo derivative in 66% yield.²³

The deprotonation of the different pyridine or pyridazine esters **11-14** was attempted using $(\text{TMP})_3\text{CdLi}$ in THF at room temperature for 2 h, and the metalated species intercepted with iodine (Table 2). Conducting the reaction from ethyl picolinate (**11**) using 0.5 equivalent of base resulted in the major formation of the 3-iodo derivative **15**, which was isolated in 58% yield (Entry 1). Ethyl isonicotinate (**12**) similarly furnished the 3-iodo compound **16**, and the yield of 65% could be slightly improved to 72% using 1 equivalent of base (Entries 2,3). Surprisingly, methyl pyridazine-4-carboxylate (**13**) behaved differently when submitted to 0.5 equiv of base, with a complex mixture of mono- and diiodides formed (Entry 4). When treated under the same conditions, ethyl nicotinate (**14**)

allowed the synthesis of the 4-iodo derivative **17** (Entry 5). The latter could not be isolated due its unstability over silica gel, but could be identified by NMR. It was involved without purification in a known copper-catalyzed reaction²⁴ with pyrazole, to provide the expected derivative **18** in a two steps 38% yield (Scheme 3).

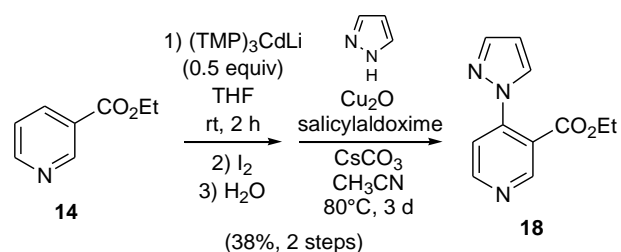
TABLE 2. Deprotonation of 11-14 using (TMP)₃CdLi Followed by Trapping with I₂.

11: ethyl picolinate
12: ethyl isonicotinate
13: methyl pyridazine-4-carboxylate
14: ethyl nicotinate

15-17

entry	substrate	x	product, yield
1	11	0.5	 15 , 58%
2	12	0.5	 16 , 65%
3	12	1	16 , 72%
4	13	0.5	mixture -
5	14	0.5	 17 , -

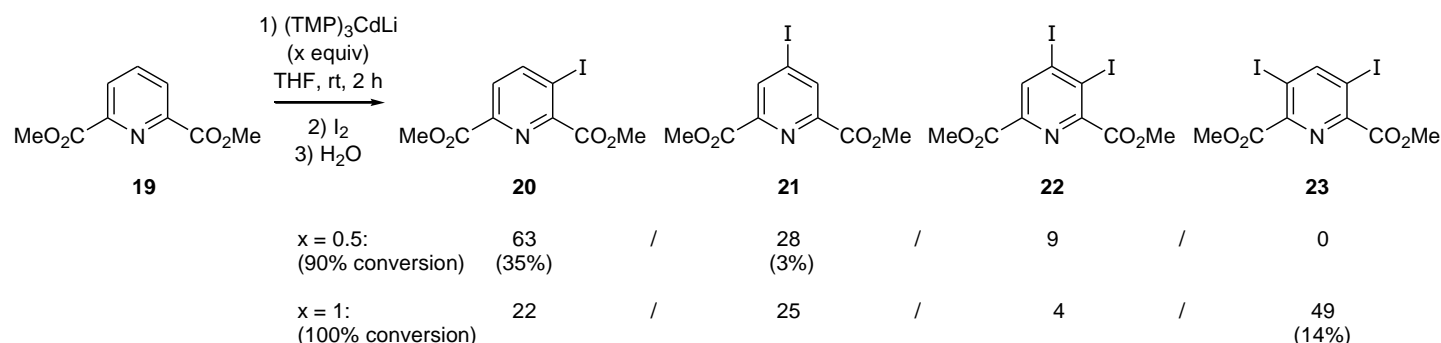
SCHEME 3. Synthesis of Ethyl 4-(pyrazol-1-yl)nicotinate (18**).**



It was then decided to involve in the deprotonation-trapping sequence methyl pyridine-2,6-dicarboxylate (**19**). By using 0.5 equivalent of (TMP)₃CdLi, the 3-iodo, 4-iodo and 3,4-diiodo derivatives **20-22** were obtained in a 63:28:9 ratio. Whereas the main compounds **20** and **21** were

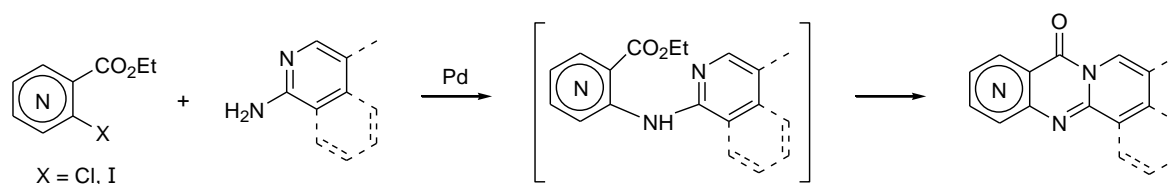
isolated from the mixture in 35 and 3% yield, respectively, methyl 3,4-diiodopyridine-2,6-dicarboxylate (**22**) was only identified from the NMR spectra of the crude. Turning to 1 equivalent of base resulted in the formation of a fourth derivative, methyl 3,5-diiodopyridine-2,6-carboxylate (**23**), together with the previous iodides. It was isolated from the 22:25:4:49 mixture of the 3-iodo, 4-iodo, 3,5-diiodo and 3,4-diiodo compounds in a modest 14% yield (Scheme 4).

SCHEME 4. Deprotonation of 19 using (TMP)₃CdLi Followed by Trapping with I₂.



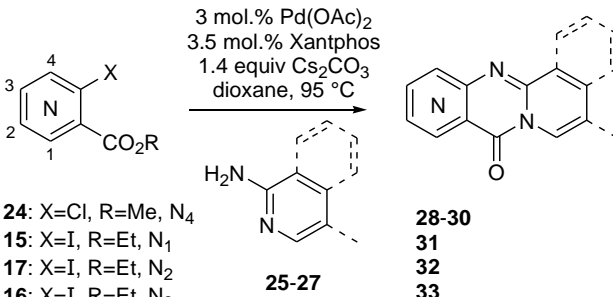
Aiming to valorize the newly synthesized ethyl iodopyridinecarboxylates **15-17**, we studied their reactivity in palladium-catalyzed cross-coupling reactions. Especially, as done previously on ethyl halogenothiophenecarboxylates,²⁵ we decided to couple those compounds, as well as methyl 2-chloronicotinate (**24**), with 2-aminopyridines **25-27** in order to access to polycyclic compounds containing a pyridopyrimidinone moiety (Scheme 5, Table 3). Indeed, the pyridopyrimidinone core is present in number of biologically active substances. For example, aza analogues of methaqualone²⁶ and 2-substituted-3-arylpyrido[2,3-*d*]pyrimidinones²⁷ proved to be anticonvulsant agents whereas some aza-quinazolinones²⁸ were described as antagonists of CXCR3. Aza-tryptanthrins exhibited antitrypanosomal activity²⁹ and inhibited *Plasmodium falciparum* cyclin-dependent kinases.³⁰

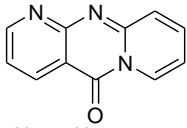
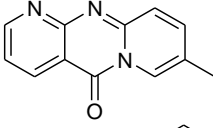
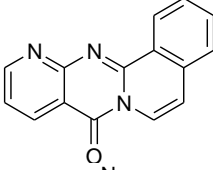
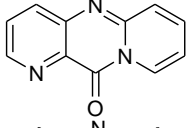
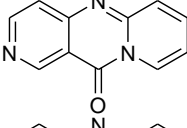
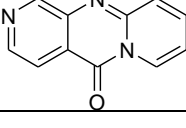
SCHEME 5. Synthetic Scheme for the Synthesis of Tricyclic (or Tetracyclic) Compounds.



Optimization of the reaction conditions was conducted by coupling methyl 2-chloronicotinate (**24**) with 2-aminopyridine (**25**). Whereas ethyl bromothiophenecarboxylates were very sensitive to the reaction conditions (several cycles of evacuation-backfilling with argon were needed) and required high catalyst loading (7 mol.% palladium acetate and 5 mol.% Xantphos), halogenopyridinecarboxylates gave good results using only 3 mol.% palladium acetate and 3.5 mol.% Xantphos. Moreover, a simple purge of argon was sufficient. The C-N coupling followed by the intramolecular cyclization involving the nitrogen atom of the pyridine ring and the carbonyl moiety of the carboxylate took place at room temperature but in a very low yield. Turning to 55 °C and 18 h of reaction allowed the formation of the expected product **28** in 20% yield. Finally, the best yield (66%) was obtained conducting the reaction at 95 °C for 24 h (Entry 1). Those conditions were then extended to the use of 2-amino-5-methylpyridine (**26**) and 1-aminoisoquinoline (**27**). The methylated compound **29** was obtained after only 2.5 h of reaction in 79% yield (Entry 2), and the tetracyclic compound **30** in a lower 43% yield (Entry 3). Involving the iodo esters **15-17** in the reaction similarly resulted in the formation of the tricyclic compounds **31-33** (Entries 4-6). Whereas **30-33** are new compounds, **28** and **29** were soon described in the literature;³¹ they were obtained thanks to a two-step process including Ullman reaction of 2-halogenonicotinic acid with 2-aminopyridine-1-oxides, and subsequent intramolecular cyclization of the resulting 3-carboxy-2,2'-bipyridylamin-1'-oxides using PCl₃.

TABLE 3. Buchwald-Hartwig Cross-coupling of 24, 15-17.

	
<p>24: X=Cl, R=Me, N₄ 15: X=I, R=Et, N₁ 17: X=I, R=Et, N₂ 16: X=I, R=Et, N₃</p>	<p>28-30 31 32 33</p>
entry	substrates product, yield

1	24 + 25		28 , 66%
2	24 + 26		29 , 79%
3	24 + 27		30 , 43%
4	15 + 25		31 , 62%
5	16 + 25		32 , 50% ^a
6	17 + 25		33 , 52%

^a For 2 steps.

Pharmacology

Applying the agar plate diffusion technique,³² the newly synthesized compounds **28-33** were screened in vitro for their bactericidal activity against Gram positive bacteria (*Staphylococcus aureus*) and Gram negative bacteria (*Escherichia Coli* and *Pseudomonas aeruginosa*), and for their fungicidal activity against *Fusarium*, *Aspergillus niger* and *Candida albicans* (Table 4). The compounds **32** and **33** showed a good bactericidal activity, similar to that of ciprofloxacin, against *Pseudomonas aeruginosa* whereas **28** and **30** showed a good fungicidal activity, similar to that of nystin, against *Fusarium*, and **32** against *Candida albicans*.

TABLE 4. Bactericidal and fungicidal activity of the compounds 28-33, and ciprofloxacin and nystin.^a

entry	compound	<i>Staphylococcus aureus</i>	<i>Escherichia coli</i>	<i>Pseudomonas aeruginosa</i>	<i>Fusarium</i>	<i>Aspergillus niger</i>	<i>Candida albicans</i>
1	28	19 (++)	-	24 (++)	27 (+++)	-	22 (++)
2	29	18 (++)	-	19 (++)	17 (++)	-	16 (++)
3	30	-	-	-	56 (++++++)	18 (++)	19 (++)
4	31	16 (++)	-	-	-	-	18 (++)

5	32	17 (++)	18 (++)	25 (+++)	20 (++)	-	25 (+++)
6	33	17 (++)	-	25 (+++)	-	-	22 (++)
7	Ciprofloxacin	+++	+++	+++			
8	Nystin				+++	+++	+++

^a The diameters of zones of inhibition are given in mm. Stock solution: 5 µg in 1 mL of DMF. 1 mL of stock solution in each hole of each paper disk. +: < 15 mm; ++: 15-24 mm; +++: 25-34 mm; ++++: 35-44 mm, etc.

The compounds **18** and **28-33** were also tested against a human liver carcinoma cell line (HEPG2), a human breast carcinoma cell line (MCF7), and a cervix carcinoma cell line (HELA) (Table 5). Moderate cytotoxic activities were observed; the compounds **18** and **28** were found to have more promising activities toward HEPG2 and MCF7, respectively, compared to a reference drug (doxorubicin).

TABLE 5. In vitro cytotoxic activity (IC₅₀) of the compounds 18, 28-33, and doxorubicin against a liver carcinoma cell line (HEPG2), a human breast carcinoma cell line (MCF7), and a cervix carcinoma cell line (HELA).^a

entry	compound	HEPG2 (µg.mL ⁻¹)	MCF7 (µg.mL ⁻¹)	HELA (µg.mL ⁻¹)
1	18	0.89	2.38	1.51
2	28	1.70	0.78	1.39
3	29	2.27	2.99	3.11
4	30	1.77	1.73	1.09
5	31	1.47	2.50	3.26
6	32	1.81	2.34	2.31
7	33	1.58	1.96	1.70
8	Doxorubicin	0.60	0.70	0.85

^a IC₅₀ is defined as the concentration which results in a 50% decrease in cell number as compared with that of the control structures in the absence of an inhibitor.

Conclusion

All pyridine nitriles and esters were metalated at the position next to the directing group using 0.5 equivalent of (TMP)₃CdLi in tetrahydrofuran at room temperature for 2 h. Subsequent trapping with iodine afforded the iodo derivatives in yields ranging from 30 to 65%. The ethyl iodopyridinecarboxylates thus obtained were then involved in a one pot palladium-catalyzed cross-coupling reaction/cyclization using 2-aminopyridine to afford new polycyclic compounds containing a

pyridopyrimidinone moiety, which were evaluated for their bactericidal and fungicidal activity. Some of the newly synthesized compounds were tested for their antitumor activity.

Because of the toxicity of cadmium compounds,³³ the use of other ate bases was before considered. Polar mixtures including alkali (or alkaline-earth metal) were ruled out because of their lack of compatibility with both reactive functions and sensitive aromatic heterocycles. Lithium aluminate and cuprate were similarly discarded, sensitive heterocycles being converted with these bases at low temperatures.^{12,15} The 1:1 LiTMP/(TMP)₂Zn lithium-zinc combination, prepared from ZnCl₂·TMEDA and 3 equivalents of LiTMP, allows efficient deprotonation reactions of aromatic substrates.³⁴ Nevertheless, it was not employed here because reactions using it are in general more weakly chemoselective,^{16b} probably due to the presence of free LiTMP. Real lithium zincates could be more suitable for the functionalization of heteroaromatic esters and nitriles; studies in order to identify bases allowing more efficient and chemoselective reactions are currently under investigation.

Experimental Section

General Procedure A (Deprotonation using 0.5 equiv CdCl₂·TMEDA and 1.5 equiv LiTMP Followed by Trapping using I₂). To a stirred cooled (0 °C) solution of 2,2,6,6-tetramethylpiperidine (0.52 mL, 3.0 mmol) in THF (5 mL) were added BuLi (1.6 M hexanes solution, 3.0 mmol) and, 5 min later, CdCl₂·TMEDA³⁵ (0.30 g, 1.0 mmol). The mixture was stirred for 10 min at 0 °C before introduction of the substrate (2.0 mmol). After 2 h at room temperature, a solution of I₂ (0.76 g, 3.0 mmol) in THF (5 mL) was added. The mixture was stirred overnight before addition of an aq saturated solution of Na₂S₂O₃ (10 mL) and extraction with EtOAc (3 x 20 mL). The combined organic layers were dried over MgSO₄, filtered and concentrated under reduced pressure.

General Procedure B (Deprotonation using 1.0 equiv CdCl₂·TMEDA and 3.0 equiv LiTMP Followed by Trapping using I₂). To a stirred cooled (0 °C) solution of 2,2,6,6-tetramethylpiperidine (1.1 mL, 6.0 mmol) in THF (5 mL) were successively added BuLi (1.6 M hexanes solution, 6.0 mmol) and, 5 min later, CdCl₂·TMEDA³⁵ (0.60 g, 2.0 mmol). The mixture was stirred for 10 min at 0 °C

before introduction of the substrate (2.0 mmol). After 2 h at room temperature, a solution of I₂ (1.5 g, 6.0 mmol) in THF (5 mL) was added. The mixture was stirred overnight before addition of an aq saturated solution of Na₂S₂O₃ (10 mL) and extraction with EtOAc (3 x 20 mL). The combined organic layers were dried over MgSO₄, filtered and concentrated under reduced pressure.

2-Cyano-3-iodopyridine (5a).^{21a} **5a** was obtained according to the general procedure A starting from 2-cyanopyridine (0.21 g), but keeping the metallation temperature at 0 °C, and was isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 80/20) as a white powder (0.18 g, 39%): mp 98 °C; ¹H NMR (200 MHz, CDCl₃) δ 7.26 (dd, 1 H, *J* = 8.2 and 4.6), 8.24 (dd, 1 H, *J* = 8.2 and 1.4), 8.68 (dd, 1 H, *J* = 4.6 and 1.4); ¹³C NMR (50 MHz, CDCl₃): δ 117.4, 127.5, 137.9, 138.5, 146.6, 149.4. HRMS: calcd for C₆H₃IN₂ (M⁺) 229.9341, found 229.9345.

2-Cyano-6-iodopyridine was identified by its ¹H NMR spectra (300 MHz, CDCl₃): δ 7.49 (t, 1 H, *J* = 7.8), 7.68 (dd, 1 H, *J* = 7.6 and 1.0), 7.95 (dd, 1 H, *J* = 7.8 and 1.0).

2-Cyano-3,4-diiodopyridine was identified by its ¹H NMR spectra (300 MHz, CDCl₃): δ 7.62 (d, 1 H, *J* = 8.4), 7.79 (d, 1 H, *J* = 8.4).

2-Cyano-3,6-diiodopyridine (5b).^{21a} **5b** was obtained according to the general procedure B starting from 2-cyanopyridine (0.21 g), and isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 90/10) as a beige powder (0.20 g, 28%): mp 140 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.62 (d, 1 H, *J* = 8.4), 7.80 (d, 1 H, *J* = 8.4); ¹³C NMR (75 MHz, CDCl₃): δ 97.7, 116.3, 116.5, 139.1, 140.0, 147.5. HRMS: calcd for C₆H₃I₂N₂ ([M+H]⁺) 356.8386, found 356.8387.

2-Cyano-3,4,6-triiodopyridine (5c). **5c** was obtained according to the general procedure B starting from 2-cyanopyridine (0.21 g), and isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 90/10) as a yellow powder (0.19 g, 20%): mp 211 °C; ¹H NMR (300 MHz, CDCl₃) δ 8.39 (s, 1 H); ¹³C NMR (75 MHz, CDCl₃): δ 113.2, 115.9, 117.0, 122.4, 139.7, 147.1. HRMS: calcd for C₆H₂I₃N₂ ([M+H]⁺) 482.7352, found 482.7352.

4-Cyano-3-iodopyridine (6a).^{21a} **6a** was obtained according to the general procedure A starting from 4-cyanopyridine (0.21 g), and isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 70/30) as a beige powder (0.20 g, 44%): mp 122 °C; ¹H NMR (200 MHz, CDCl₃) δ 7.52 (dd, 1 H, *J* = 4.8 and 0.6), 8.71 (d, 1 H, *J* = 5.0), 9.10 (s, 1 H); ¹³C NMR (75 MHz, CDCl₃): δ 96.4, 117.0, 127.1, 127.9, 148.9, 158.0. HRMS: calcd for C₆H₃IN₂ (M⁺) 229.9341, found 229.9345.

4-Cyano-3,5-diiodopyridine (6b).^{21a} **6b** was obtained according to the general procedure A starting from 4-cyanopyridine (0.21 g), but keeping the metallation temperature at 0 °C, and was isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 90/10) as a beige powder (0.14 g, 20%): mp 153 °C; ¹H NMR (300 MHz, CDCl₃) δ 8.97 (s, 2 H); ¹³C NMR (75 MHz, CDCl₃): δ 97.3 (2C), 118.4, 134.5, 156.4 (2C). HRMS: calcd for C₆H₂I₂N₂Na ([M+Na]⁺) 378.8205, found 378.8207.

4-Cyano-2-iodopyridine (6c).³⁶ **6c** was obtained according to the general procedure A starting from 4-cyanopyridine (0.21 g), and isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 70/30) as a beige powder (46 mg, 10%): mp 76 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.50 (dd, 1 H, *J* = 5.0 and 1.4), 7.96 (t, 1 H, *J* = 1.4), 8.56 (dd, 1 H, *J* = 5.0 and 1.4); ¹³C NMR (75 MHz, CDCl₃): δ 114.6, 117.8, 121.5, 124.1, 136.0, 151.3. HRMS: calcd for C₆H₃IN₂ (M⁺) 229.9341, found 229.9345.

4-Cyano-2,3-diiodopyridine (6d). **6d** was obtained according to the general procedure B starting from 4-cyanopyridine (0.21 g), and isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 80/20) as a beige powder (0.36 g, 51%): mp 215 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.84 (d, 1 H, *J* = 5.3), 8.08 (d, 1 H, *J* = 5.3); ¹³C NMR (75 MHz, CDCl₃): δ 110.6, 119.1, 121.0, 127.1, 133.1, 151.7. HRMS: calcd for C₆H₂I₂N₂ (M⁺) 355.8307, found 355.8341.

3-Cyano-2-iodopyridine (7). **7** was obtained according to the general procedure A starting from 3-cyanopyridine (0.21 g), and isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 80/20) as a beige powder (0.28 g, 61%): mp 127 °C; ¹H NMR (200 MHz, CDCl₃) δ

7.43 (dd, 1 H, $J = 7.7$ and 4.9), 7.82 (dd, 1 H, $J = 7.7$ and 2.0), 8.54 (dd, 1 H, $J = 4.9$ and 2.0); ^{13}C NMR (50 MHz, CDCl_3): δ 117.7, 119.9, 121.0, 122.5, 141.2, 152.9. HRMS: calcd for $\text{C}_6\text{H}_3\text{IN}_2$ (M^+) 229.9341, found 229.9345. These data are analogous to those previously described.¹⁹

2-Cyano-3-iodopyrazine (8).³⁷ **8** was obtained according to the general procedure A starting from cyanopyrazine (0.21 g), and isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 90/10) as a yellow powder (0.20 g, 43%): mp 107 °C; ^1H NMR (300 MHz, CDCl_3) δ 8.54 (d, 1 H, $J = 2.4$), 8.65 (d, 1 H, $J = 2.4$); ^{13}C NMR (75 MHz, CDCl_3): δ 116.3, 120.7, 138.2, 143.2, 147.2. HRMS: calcd for $\text{C}_5\text{H}_2\text{IN}_3\text{Na}$ ($[\text{M}+\text{Na}]^+$) 253.9191, found 253.9192.

Ethyl 5-iodothiophene-2-carboxylate (10). **10** was obtained according to the general procedure A starting from ethyl thiophene-2-carboxylate (0.31 g), and isolated after purification by flash chromatography on silica gel (eluent: heptane/ CH_2Cl_2 90/10) as a yellow oil (0.43 g, 77%): ^1H NMR (300 MHz, CDCl_3) δ 1.36 (t, 3 H, $J = 7.1$), 4.33 (q, 2 H, $J = 7.1$), 7.25 (d, 1 H, $J = 3.9$), 7.42 (d, 1 H, $J = 3.9$). These data are similar to those described.²² ^{13}C NMR (75 MHz, CDCl_3): δ 14.4, 61.5, 82.6, 134.4, 137.8, 139.8, 161.0.

Ethyl 3-iodopicolinate (15).³⁸ **15** was obtained according to the general procedure A starting from ethyl picolinate (0.30 g), and isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 80/20) as a yellow oil (0.32 g, 58%): ^1H NMR (200 MHz, CDCl_3) δ 1.45 (t, 3 H, $J = 7.1$), 4.47 (q, 2 H, $J = 7.1$), 7.11 (dd, 1 H, $J = 8.2$ and 4.7), 8.25 (dd, 1 H, $J = 8.2$ and 1.4), 8.62 (dd, 1 H, $J = 4.7$ and 1.4); ^{13}C NMR (50 MHz, CDCl_3): δ 14.1, 62.1, 92.1, 126.1, 126.2, 148.2, 152.3, 165.7. HRMS: calcd for $\text{C}_8\text{H}_8\text{INO}_2$ (M^+) 276.9600, found 276.9609.

Ethyl 3-iodoisonicotinate (16).²³ **16** was obtained according to the general procedure A starting from ethyl isonicotinate (0.30 g), and isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 80/20) as an orange oil (0.36 g, 65%): ^1H NMR (200 MHz, CDCl_3) δ 1.41 (t, 3 H, $J = 7.1$), 4.42 (q, 2 H, $J = 7.1$), 7.62 (d, 1 H, $J = 4.9$), 8.60 (d, 1 H, $J = 4.8$), 9.08 (s, 1H); ^{13}C NMR (50

MHz, CDCl₃): δ 14.0, 62.4, 92.3, 124.3, 142.3, 149.0, 159.4, 164.9. HRMS: calcd for C₈H₈INO₂ (M⁺) 276.9600, found 276.9609.

Methyl 3-iodopyridazine-4-carboxylate. A pure fraction was isolated (eluent: heptane/EtOAc 85/15) from the crude obtained according to the general procedure A as a yellow solid (degradation to a dark residue upon standing): ¹H NMR (300 MHz, CDCl₃): δ 7.66 (d, 1 H, *J* = 5.0), 9.26 (d, 1 H, *J* = 5.0), 4.01 (s, 3 H); ¹³C NMR (75 MHz, CDCl₃): δ 53.7, 77.4, 126.2, 135.3, 150.6, 164.4.

Methyl 3,5-diiodopyridazine-4-carboxylate. A pure fraction was isolated (eluent: heptane/EtOAc 85/15) from the crude obtained according to the general procedure A as a yellow solid (degradation to a dark residue upon standing): ¹H NMR (300 MHz, CDCl₃): δ 9.41 (s, 1 H), 4.03 (s, 3 H); ¹³C NMR (75 MHz, CDCl₃): δ 54.1, 97.4, 120.2, 145.6, 157.9, 165.4.

Ethyl 4-iodonicotinate (17). **17** was obtained according to the general procedure A starting from ethyl nicotinate (0.30 g), but could not be purified by flash chromatography on silica gel because of its low stability. It was identified by NMR: ¹H NMR (200 MHz, CDCl₃) δ 1.42 (t, 3 H, *J* = 7.1), 4.14 (q, 2 H, *J* = 7.1), 7.94 (d, 1 H, *J* = 5.3), 8.23 (d, 1 H, *J* = 5.3), 8.93 (s, 1 H). ¹³C NMR (50 MHz, CDCl₃): δ 14.1, 62.1, 106.1, 131.0, 136.2, 151.0, 152.0, 164.7. The crude was directly involved in the reactions giving the compounds **18** and **32**.

Ethyl 4-(pyrazol-1-yl)nicotinate (18). **18** was obtained from the crude compound **17** by adapting a procedure described,²⁴ and was isolated after purification by flash chromatography on silica gel (eluent: heptane/EtOAc 70/30) as an orange oil (0.17 g, 2 steps, 38%): ¹H NMR (300 MHz, CDCl₃) δ 1.24 (t, 3 H, *J* = 7.1), 4.31 (q, 2 H, *J* = 7.2), 6.50 (dd, 1 H, *J* = 2.4 and 1.8), 7.48 (d, 1 H, *J* = 5.4), 7.75 (d, 1 H, *J* = 1.5), 7.83 (d, 1 H, *J* = 2.7), 8.75 (d, 1 H, *J* = 5.1), 8.92 (s, 1H); ¹³C NMR (75 MHz, CDCl₃): δ 14.1, 62.1, 108.8, 117.1, 121.9, 129.5, 142.6, 145.1, 151.5, 152.9, 165.9. HRMS: calcd for C₁₁H₁₂N₃O₂ ([M+H]⁺) 218.0930, found 218.0932.

Methyl 3-iodopyridine-2,6-dicarboxylate (20). **20** was obtained according to the general procedure A starting from methyl pyridine-2,6-dicarboxylate (0.39 g), and isolated by flash chromatography on silica gel (eluent: heptane/CH₂Cl₂ 80/20) as a pale orange powder (0.22 g, 35%): mp 88–90 °C; ¹H NMR (300 MHz, CDCl₃) δ 4.00 (s, 3 H), 4.01 (s, 3 H), 7.89 (d, 1 H, *J* = 8.1), 8.42 (d, 1 H, *J* = 8.1); ¹³C NMR (50 MHz, CDCl₃): δ 53.3 (2C), 95.3, 127.1, 146.8, 149.6, 152.6, 164.7, 165.5. HRMS: calcd for C₉H₈INO₄ (M⁺) 320.9498, found 320.9496.

Methyl 4-iodopyridine-2,6-dicarboxylate (21). **21** was obtained according to the general procedure A starting from methyl pyridine-2,6-dicarboxylate (0.39 g), and isolated by flash chromatography on silica gel (eluent: heptane/CH₂Cl₂ 80/20) in 3% (18 mg) yield: ¹H NMR (300 MHz, CDCl₃) δ 4.02 (s, 6H), 8.66 (s, 2H). ¹³C NMR (50 MHz, CDCl₃): δ 53.2 (2C), 106.8, 136.9 (2C), 148.0 (2C), 163.6 (2C). These data are similar to those previously described.³⁹ HRMS: calcd for C₇H₆INO₂ [(M-C₂H₂O₂)⁺] 262.9443, found 262.9469.

Methyl 3,4-diiodopyridine-2,6-dicarboxylate (22). **22** formed using the general procedure A and B, and was identified by its NMR data: ¹H NMR (300 MHz, CDCl₃) δ 3.98 (s, 6H), 8.88 (s, 1H). HRMS: calcd for C₈H₅I₂NO₃ [(M-CH₂O)⁺] and C₇H₅I₂NO₂ [(M-C₂H₂O₂)⁺] 416.8359 and 388.8410, found 416.8379 and 388.8426.

Methyl 3,5-diiodopyridine-2,6-dicarboxylate (23). **23** was obtained according to the general procedure B starting from methyl pyridine-2,6-dicarboxylate (0.39 g), and isolated by flash chromatography on silica gel (eluent: heptane/CH₂Cl₂ 80/20) as a beige powder (0.13 g, 14%): mp 128–130 °C; ¹H NMR (300 MHz, CDCl₃) δ 3.99 (s, 6 H), 8.88 (s, 1 H); ¹³C NMR (50 MHz, CDCl₃): δ 53.4 (2C), 93.4 (2C), 150.6, 159.5, 164.9 (2C). HRMS: calcd for C₉H₇I₂NO₄ (M⁺) 446.8465, found 446.8447.

General Procedure C for Buchwald-Hartwig Cross-coupling. A solution of Pd(OAc)₂ (10 mg, 3 mol%), Xantphos (30 mg, 3.5 mol%) and Cs₂CO₃ (675 mg, 1.4 equiv) was prepared under argon in dioxane. When the temperature reached 55 °C, the appropriate halogenopyridine (1.5 mmol, 1 equiv)

was added under argon and then 5 to 10 minutes later (temperature about 80 °C), the aminopyridine (1.8 mmol, 1.2 equiv) was finally introduced. The reaction mixture was stirred at 95 °C for 2.5 h to 24 h under argon (reaction was followed by thin layer chromatography). After cooling to room temperature, the reaction mixture was filtered, and the cake was washed with EtOAc. The filtrate was concentrated under reduced pressure.

Dipyrido[1,2-*a*:2',3'-*d*]pyrimidin-5-one (28). **28** was obtained according to the general procedure C starting from methyl 2-chloronicotinate (0.26 g) and 2-aminopyridine (0.17 g), and was isolated after purification by chromatography on silica gel (CHCl₃ as eluent) as a yellow solid (0.20 g, 66%): mp 220-221 °C (lit.⁴⁰ 223 °C); ¹H NMR (250 MHz, CDCl₃) δ 6.99 (m, 1 H), 7.43 (dd, 1 H, *J* = 8.0 and 4.4), 7.64-7.69 (m, 2H), 8.78 (dd, 1 H, *J* = 8.0 and 2.1), 8.89 (m, 1 H), 9.12 (dd, 1 H, *J* = 4.4 and 2.1); ¹³C NMR (62.5 MHz, CDCl₃) δ 111.3, 113.5, 120.6, 126.7, 127.0, 135.7, 137.0, 149.8, 157.4, 157.8, 159.7; IR (KBr) ν 1698, 1641, 1593, 1543, 1526, 1411 cm⁻¹; HRMS: calcd for C₁₁H₈N₃O ([M+H]⁺) 198.0662, found 198.0668.

8-Methyldipyrido[1,2-*a*:2',3'-*d*]pyrimidin-5-one (29). **29** was obtained according to the general procedure C starting from methyl 2-chloronicotinate (0.26 g) and 2-amino-5-methylpyridine (0.19 g), and was isolated after purification by chromatography on silica gel (CHCl₃ as eluent) as a yellow solid (0.25 g, 79%): mp 201-202 °C (lit.⁴⁰ 203 °C); ¹H NMR (250 MHz, CDCl₃) δ 2.48 (s, 3 H), 6.82 (dd, 1 H, *J* = 7.5 and 1.8), 7.38 (dd, 1 H, *J* = 8.0 and 4.4), 7.45 (br s, 1 H), 8.74 (dd, 1 H, *J* = 8.0 and 2.1), 8.79 (d, 1 H, *J* = 7.5), 9.08 (dd, 1 H, *J* = 4.4 and 2.1); ¹³C NMR (62.5 MHz, CDCl₃) δ 21.6, 110.9, 116.5, 120.1, 124.4, 126.0, 137.0, 147.8, 149.9, 157.7, 157.8, 159.8; IR (KBr) ν 1693, 1651, 1592, 1543, 1412 cm⁻¹; HRMS: calcd for C₁₂H₁₀N₃O ([M+H]⁺) 212.0818, found 212.0826.

Pyrido[2',3':4,5]pyrimidino[2,1-*a*]isoquinolin-8-one (30). **30** was obtained according to the general procedure C starting from methyl 2-chloronicotinate (0.26 g) and 1-aminoisoquinoline (0.26 g), and was isolated after purification by chromatography on silica gel (eluent: EtOAc/C₆H₁₂ 70/30) as a yellow solid (95 mg, 43%): mp 242-244 °C; ¹H NMR (250 MHz, CDCl₃) δ 7.17 (d, 1 H, *J* = 7.8), 7.48 (dd, 1

H, $J = 8.0$ and 4.5), 7.70-7.83 (m, 3 H), 8.67 (d, 1 H, $J = 7.8$), 8.81 (dd, 1 H, $J = 8.0$ and 2.1), 9.13 (dd, 1 H, $J = 4.5$ and 2.1); 9.30 (d, 1 H, $J = 8.0$); ^{13}C NMR (62.5 MHz, CDCl_3) δ 112.8, 114.2, 121.2, 121.5, 126.5, 127.0, 128.2, 128.9, 133.1, 133.4, 137.0, 149.1, 156.9, 157.2, 160.1; IR (KBr) ν 1679, 1645, 1593, 1555, 1423 cm^{-1} ; HRMS: calcd for $\text{C}_{15}\text{H}_{10}\text{N}_3\text{O}$ ($[\text{M}+\text{H}]^+$) 248.0818, found 248.0808.

Dipyrido[1,2-*a*:3',2'-*d*]pyrimidin-11-one (31). **31** was obtained according to the general procedure C starting from ethyl 3-iodopicolinate (0.42 g) and 2-aminopyridine (0.17 g), and was isolated after purification by chromatography on silica gel (CHCl_3 as eluent) as a yellow solid (0.12 g, 62%): mp 211 $^{\circ}\text{C}$; ^1H NMR (250 MHz, CDCl_3) δ 6.94-7.00 (m, 1 H), 7.55-7.67 (m, 2 H), 7.75 (dd, 1 H, $J = 8.5$ and 4.1), 8.14 (dd, 1 H, $J = 8.5$ and 1.5), 8.91 (dd, 1 H, $J = 4.1$ and 1.5), 9.00-9.04 (m, 1H); ^{13}C NMR (62.5 MHz, CDCl_3) δ 113.1, 126.3, 127.4, 129.0, 132.5, 135.1, 135.3, 145.6, 148.2, 148.9, 157.7; IR (KBr) ν 1702, 1641, 1543, 1519, 1469, 1413 cm^{-1} ; HRMS: calcd for $\text{C}_{11}\text{H}_8\text{N}_3\text{O}$ ($[\text{M}+\text{H}]^+$) 198.0662, found 198.0662.

Dipyrido[1,2-*a*:4',3'-*d*]pyrimidin-11-one (32). **32** was obtained according to the general procedure C starting from crude ethyl 4-iodonicotinate (general procedure A) and 2-aminopyridine (0.23 g), and was isolated after purification by chromatography on silica gel (eluent: $\text{CH}_2\text{Cl}_2/\text{Et}_3\text{N}$ 98/2) as a brown solid (0.20 g, 50% for 2 steps): mp 130 $^{\circ}\text{C}$; ^1H NMR (300 MHz, CDCl_3) δ 6.99 (m, 1H), 7.54 (m, 2H), 7.68 (ddd, 1H, $J = 9.2$, 6.5 and 1.6), 8.80 (d, 1H, $J = 6.0$), 8.93 (m, 1H), 9.64 (s, 1H). ^{13}C NMR (75 MHz, CDCl_3) δ 111.9, 113.7, 119.7, 126.4, 127.2, 139.4, 151.1, 152.0, 152.7, 152.9, 158.2. HRMS: calcd for $\text{C}_{11}\text{H}_7\text{N}_3\text{O}$ ($[\text{M}+\text{H}]^+$) 198.0667, found 198.0670.

Dipyrido[1,2-*a*:3',4'-*d*]pyrimidin-5-one (33). **33** was obtained according to the general procedure C starting from ethyl 3-iodoisonicotinate (0.42 g) and 2-aminopyridine (0.17 g), and was isolated after purification by chromatography on silica gel (eluent: $\text{CH}_2\text{Cl}_2/\text{Et}_3\text{N}$ 98/2) as an orange solid (0.15 g, 52%): mp 164 $^{\circ}\text{C}$; ^1H NMR (300 MHz, CDCl_3) δ 6.96 (m, 1H), 7.59 (m, 2H), 8.14 (dd, 1H, $J = 5.4$ and 0.7), 8.62 (br d, 1H, $J = 5.4$), 8.86 (dt, 1H, $J = 7.4$ and 1.2), 9.25 (br s, 1H). ^{13}C NMR (75 MHz, CDCl_3)

δ 113.8, 118.6, 120.2, 126.8, 127.0, 135.1, 143.1, 143.7, 149.1, 151.9, 158.3. HRMS: calcd for $C_{11}H_7N_3O$ ($[M+H]^+$) 198.0667, found 198.0672.

Pharmacology. Applying the agar plate diffusion technique,³² the compounds were screened in vitro for their bactericidal activity against Gram positive bacteria (*Staphylococcus aureus*) and Gram negative bacteria (*Escherichia Coli* and *Pseudomonas aeroginosa*), and for their fungicidal activity against *Fusarium*, *Aspergillus niger* and *Candida albicans*. In this method, a standard 5 mm diameter sterilized filter paper disc impregnated with the compound (0.3 mg/0.1 ml of DMF) was placed on an agar plate seeded with the test organism. The plates were incubated for 24 hours at 37 °C for bacteria and 28 °C for fungi. The zone of inhibition of bacterial and fungi growth around the disc was observed.

The compounds were tested against a liver carcinoma cell line (HEPG2), a human breast carcinoma cell line (MCF7), and a cervix carcinoma cell line (HELA). The method applied is similar to that reported by Skehan et al.⁴¹ using 20 Sulfo-Rhodamine-B stain (SRB). Cells were plated in 96-multiwell plate (104 cells/well) for 24 h before treatment with the test compound to allow attachment of cell to the wall of the plate. Different concentrations of the compound under test (0, 1.0, 2.5, 5.0, and 10 μ g/ml) were added to the cell monolayer in triplicate wells individual dose, and monolayer cells were incubated with the compounds for 48 h at 37°C and in atmosphere of 5% CO₂. After 48 h, cells were fixed, washed and stained with SRB stain, excess stain was washed with acetic acid and attached stain was recovered with Tris-EDTA buffer. Color intensity was measured in an ELISA reader, and the relation between surviving fraction and drug concentration is plotted to get the survival curve of each tumor cell line after the specified compound and the IC₅₀ was calculated.

Acknowledgment. We are grateful to Région Bretagne (France) and to MESRS (Algeria) for financial support to G. B, and to MESR (France) for financial support to T. T. N. We thank Rennes Métropole. We are also grateful to Faculty of Women, Ain Shams University, and National Institute of Cancer, Cairo University (Egypt) for pharmacological measurements.

Supporting Information Available: General procedures and copies of the ^1H and ^{13}C NMR spectra for compounds **5a-c**, **6a-d**, **7**, **8**, **10**, **15-18**, **20**, **23**, **28-33**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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